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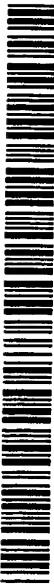
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**WO 02/06503 A2**

(54) Title: PROCESS FOR GENERATION OF HYDROGEN GAS FROM VARIOUS FEEDSTOCKS USING THERMOPHILIC BACTERIA

(57) Abstract: A method for producing hydrogen gas is provided comprising selecting a bacteria from the Order Thermotogales, subjecting the bacteria to a feedstock and to a suitable growth environment having an oxygen concentration below the oxygen concentration of water in equilibrium with air, and maintaining the environment at the predetermined pH and at a temperature of at least approximately 45°C for a time sufficient to allow the bacteria to metabolize the feedstock.

PROCESS FOR GENERATION OF HYDROGEN  
GAS FROM VARIOUS FEEDSTOCKS USING  
THERMOPHILIC BACTERIA

CONTRACTUAL ORIGIN OF THE INVENTION

The United States Government has rights in this invention pursuant to an employment relationship between the inventor and the United States Department of Energy.

BACKGROUND OF THE INVENTION

5 1. Field of the Invention

This invention relates to a method for using bacteria to produce hydrogen gas, and more particularly this invention relates to a method for using thermophilic bacteria to generate hydrogen gas from a wide variety of feedstocks.

10 2. Background of the Invention

Hydrogen gas holds promise as the fuel of the future. The U.S. currently consumes 3.6 trillion cubic feet (TCF) of hydrogen gas annually, with a worldwide consumption of about three times that amount. The U.S. demand alone is expected to increase by 40% to 5.0 TCF in the next five years.

15 Most of the hydrogen gas produced throughout the world is made from synthesis gas generated either by reformation of natural gas or from the gasification of coal. Not only are these processes costly, but they are hostile to the environment. Furthermore, these methods use fossil fuels, which are non-renewable.

Biological processes have been used previously to generate hydrogen gas.

Biological processes are particularly attractive because renewable feedstocks (i.e., biomass, even organic waste streams) are utilized. Research in this area has concentrated on three different approaches:

- 1) Using photosynthetic organisms to split water;
- 5 2) Using fermentative bacteria to digest hydrocarbons;
- 3) Using a combination of bacteria in which some of the bacteria digest the complex hydrocarbons to make an appropriate feedstock for the hydrogen-producing bacteria.

However, and as discussed more fully, infra, these efforts have not yet resulted 10 in commercial success because of one or more problems. In some cases, the metabolic processes that produce hydrogen gas are end-product or by-product inhibited, while in other cases the growth rate of the bacteria is end-product or by-product inhibited. Still in other cases, the range of suitable feedstocks is narrow, or the production of undesirable gases is high. In some processes, the rate of hydrogen gas production is 15 very low, while in all cases faster rates are highly desirable.

For example, photosynthetic processes exist (U.S. Patent No. 5,804,424 to Kaplan et al; Patent No. 4,921,800 to Vatsala, and Patent No. 4,919,813 to Weaver) whereby photosynthetic organisms (such as algae or microalgae) use light to convert water into hydrogen gas and oxygen gas. However, the yields of hydrogen gas are very 20 low in these instances because one of the end products of the reaction, oxygen gas ( $O_2$ ), irreversibly inhibits the hydrogenase enzyme responsible for hydrogen gas production. Even a very small amount of oxygen gas produced as a byproduct is sufficient to shut down the entire hydrogen gas production system by inactivating the hydrogenase enzyme. Thus, the amount of hydrogen gas produced is minimal, rarely 25 exceeding 10 ppm. Efforts to prevent this oxygen gas inhibition by modification of the hydrogenase enzyme have met with only

modest success. These altered hydrogenases can only tolerate 2% oxygen gas concentration.

Pond scum has been utilized recently to produce hydrogen gas at a rate of 3 30 ml/l-hour. A. Melis, et al., "Sustained Photobiological Hydrogen Gas Production Upon Reversible Inactivation of Oxygen Evolution in the Green Alga," *Proceedings of the 1999 U.S. DOE Hydrogen Program Review*, pp. 1-19. However, this was accomplished only while maintaining an oxygen-free environment for the algae and only for short time spans up to four days.

Another approach to producing hydrogen gas is the process of fermentation. Processes utilizing monocultures of various mesophilic bacteria produce hydrogen gas as a byproduct of anaerobic fermentative degradation of simple sugars (Roychowdhury, 4,480,035) or a combination of either formic acid or a formate plus a nitrogen source 5 (Sanford, 5,834,264). Alternatively, glucose or glucose-containing polysaccharide feedstocks are utilized (Taguchi, 5,350,692). These processes confer advantages over photosynthetic processes because fermentative processes use less water, require no direct input of solar energy, and eliminate the need for a container with a large translucent surface area. Fermentative mesophilic bacteria only need an input stream 10 consisting of an appropriate aqueous medium and substrate and an output stream to remove the generated waste products and gases. Unfortunately, monocultures of mesophilic bacteria may be easily contaminated. This is a major drawback in an industrial-scale fermenter where complete sterilization is difficult and where contamination is almost unavoidable.

15 A variation of the fermentation approach has been to co-culture a number of different types of bacteria, where the net effect is to produce hydrogen gas as a by-product of fermentation of various carbohydrates or even sludge. (Ueno, 5,464,539). Co-culture with photosynthetic organisms has also been described. (Weaver, 4,919,813). Some of these processes produce noxious gases, such as hydrogen sulfide 20 ( $H_2S$ ) and methane ( $CH_4$ ), along with other gases that would need to be separated from the hydrogen. Composting (i.e., partial decomposition) may be required as a time-consuming preliminary step to predigest the initial feedstock and form more simple compounds that can then be utilized to produce hydrogen.

Attempts have been made to utilize mesophilic and thermophilic bacteria in 25 hydrogen gas production processes. (See for example, M.W.W. Adams, CHEMTECH, November 1991, pp. 692-699.) Such bacteria as *Pyrococcus furiosus*, *Pyrodictium brockii* and *Thermotoga maritima* were examined. However, the hydrogenase system of the *P. brockii* and *T. maritima* showed a preference for  $H_2$  oxidation, versus  $H_2$  evolution. Also, the thermostability of *T. maritima* enzyme appeared inferior vis-a-vis *P. 30 furiosus*.

A need exists in the art for a biological process for generating hydrogen gas on an industrial scale. The process should utilize a wide variety of hydrocarbon sources. Furthermore, the process should maximize hydrogen gas production and minimize hydrogen sulfide gas production. The process should have an inherent feature for

minimizing its contamination.

#### SUMMARY OF THE INVENTION

An object of the present invention is to provide a process for producing hydrogen that overcomes many of the disadvantages of the prior art.

- 5        Another object of the present invention is to provide a robust catabolic process for producing hydrogen wherein a single species or several species from a single Order are cultured. A feature of the invention is that the organism is eurytolerant. An advantage of the invention is that stringent environmental controls such as air-tight environs, stringent levels of nutrients and media, and predigestion, are not required.
- 10      Another advantage of the process is that hydrogen is produced with minimal production of nitrous oxides ( $\text{NO}_x$ ), sulfur oxides ( $\text{SO}_x$ ), CO and  $\text{CH}_4$ , all of which are typically generated when hydrogen is produced via hydrocarbon gasification.

Yet another object of the present invention is to provide a process for producing hydrogen using thermophilic bacteria. A feature of the invention is that the process can utilize a variety of hydrocarbons as its feedstock. Other features of the invention are that the pH of the bioreaction is controlled, that the concentrations of potential sulfur-donors in the feedstock and aqueous medium are controlled, and that measured and limited amounts of oxygen gas are introduced into the process which the thermophilic organisms can either use or tolerate as facultative anaerobes and which oxygen gas may reduce the tendency to form hydrogen sulfide. An advantage of this invention is that hydrogen production is maximized while the production of hydrogen sulfide gas is minimized.

- Still another object of the present invention is to provide a method of waste treatment that is also an environmentally-friendly process for producing hydrogen gas.
- 25      A feature of the invention is that bacteria used in this process are omnivorous, and as such, the feedstock may consist of items from any one of a number of classes of complex hydrocarbons: such as starch, any one of a number of sugars, xylans, and/or celluloses, any one of a number of amino acids, long chain fatty acids, proteins, oils (including petroleum products) and combinations thereof. An advantage of this process is that the organisms incorporated in the process can utilize very complex hydrocarbons without requiring pre-processing to produce simple sugars or their catabolites. Therefore, the process is well suited to convert a wide variety of waste streams and biomass into hydrogen gas.
  - 30      Another object of the present invention is to provide a controllable biological process for producing hydrogen gas from a wide variety of complex hydrocarbons; at

- higher rates per unit volume; with reduced sensitivity to the presence of O<sub>2</sub>; with little sensitivity to the generated H<sub>2</sub>; with minimal interference from other organisms; plus a low production of CO<sub>2</sub> and H<sub>2</sub>S. A feature of this invention is that it operates using high temperatures, which reduce the risk of methane production and hydrogen sulfide
- 5 production by other types of bacteria. An advantage of this process is that environmentally-unfriendly production of methane gas and hydrogen sulfide gas is minimized without resorting to composting the stock materials, without excessively limiting the range of feedstocks, and without resorting to other procedures to reduce or eliminate methanogenic bacteria and sulfur-reducing bacteria or their activities.
- 10 Because few other organisms are capable of growth or even life at these temperatures, maintaining clean cultures, even in industrial-scale operations, is more feasible. Also, the desired thermophilic organisms remain viable for months at room temperature, but only grow when the temperature is elevated. Therefore, control and containment of the organism is simplified.
- 15 Still another object of the present invention is to provide a process which uses bacteria of the Order *Thermotogales*, to produce hydrogen gas in useful quantities. A feature of the invention is that the process utilizes physiological buffers and titration to maintain a pH in the reaction environment. An advantage of the process is that hydrogen can be continuously produced at rates above about 5 ml/l-hour.
- 20 Still another object of the present invention is to provide a biological process for producing hydrogen gas wherein the process utilizes facultative anaerobic bacteria which are also thermophilic. A feature of the invention is that the process can tolerate and may even use oxygen. An advantage of the process is its applicability to industrial situations. Another advantage is that the presence of oxygen may reduce the formation
- 25 of H<sub>2</sub>S concomitantly generated.
- Yet another object of the present invention is to provide a process for producing hydrogen gas that operates at subpyrolytic temperatures. A feature of the invention is that the process uses catabolism of hydrocarbons to produce hydrogen gas and carbon dioxide in a ratio of nearly 2:1. An advantage of the process is that unlike thermal
- 30 processes, very little NO<sub>x</sub>, SO<sub>x</sub>, CO and CH<sub>4</sub> is produced.

Briefly, a process for producing H<sub>2</sub> is provided comprising confining bacteria from the Order *Thermotogales* to an environment suitable for allowing the bacteria to undergo metabolism; maintaining the environment at an oxygen concentration below the oxygen concentration of water in equilibrium with air; and removing hydrogen from the

environment during the metabolism.

The invention also provides for a method for producing H<sub>2</sub> comprising selecting a bacteria from the Order *Thermotogales*; subjecting the bacteria to an environment containing a feedstock and a media, wherein the media contains oxygen gas in a

- 5 concentration below the oxygen concentration of water in equilibrium with air; and maintaining the environment at a temperature, pressure and pH sufficient to allow the bacteria to metabolize the feedstock.

Additional objects, features and advantages are found throughout the specification or will become apparent to those skilled in the art upon examination of the

- 10 description that follows. Other objects may be learned by the practice of this invention.

#### DETAILED DESCRIPTION OF THE INVENTION

Instead of using algae, mesophilic bacteria or co-cultures of bacteria, thermophilic bacteria of the Order *Thermotogales* are utilized for the production of hydrogen gas (H<sub>2</sub>). The inventors have determined experimentally that bacteria of the

- 15 Order *Thermotogales*, previously thought to be viable only in an anaerobic fermentative environment, can be used to produce hydrogen gas in a process that can tolerate relatively high concentrations of oxygen without apparent loss of hydrogen-producing ability. Some of these types of organisms can even tolerate oxygen levels very close to that of ambient air for short periods of time at room temperature.

20 In light of the forgoing, surprisingly and unexpectedly, the inventors have found that their invented process is not a true fermentative process, despite wide-spread claims in the scientific literature that incorrectly label the Order *Thermotogales* as obligate (true) anaerobes. Thus, in industrial-scale systems, small leaks in the fermentation apparatus will not upset or kill the culture.

25 The inventors speculate that any residual oxygen gas that is present combines with carbon atoms as the hydrocarbon feedstock is metabolized, thereby minimizing the tendency to form sulfides from the sulfates in solution. Moreover, the presence of oxygen gas in this process may reduce the formation of H<sub>2</sub>S by any processes that could create it. The invented process illustrates that each of the members of the Order

30 *Thermotogales* can be utilized with specific feedstocks to optimize yield, so that while one particular Species may produce more hydrogen when the feedstock is a simple sugar, others may prefer more complex carbohydrates, proteins or oils. Co-cultures of complimentary Species may act in concert to completely degrade hard-to-utilize waste materials to produce hydrogen gas. Experimental data reveals that oxygen

concentrations in the process liquor actually increases in some situations. Furthermore, and as noted elsewhere in this specification, the invented protocol generates hydrogen gas ( $H_2$ ) at a 2:1 ratio with  $CO_2$ , versus the expected 1:1 ratio. Not only is the bacteria utilizing the hydrocarbon feedstock to generate hydrogen, but the process also may be  
5 allowing the microbes to split water to produce the unexpected oxygen and hydrogen.

Thermophilic (heat-loving) bacteria become active in a temperature range that is  
much higher than for most other organisms. There is no one temperature range that is applicable to all thermophilic organisms. This is particularly true for members of the  
10 Order *Thermotogales*. Below a certain temperature, these organisms change form and appear coccoid (spherical). At higher temperatures these rod-shaped organisms metabolize, and at some temperature begin reproduction. After reaching a peak in organism growth at an optimal temperature, organism growth decreases with further increases in temperature. The range of temperatures over which an organism is seen  
15 to grow is referred to for the purposes of this document as the "permissive range" for that particular Species. The permissive range and the optimum temperature differs for each species within the Order *Thermotogales*.

Bacteria of the Order *Thermotogales* can be easily maintained as a monoculture and readily inactivated and reactivated after short-term storage at room temperature.  
20 When the reaction temperature is elevated to the permissive range, the organisms readily grow and produce hydrogen gas. At this elevated temperature, most other bacteria are not capable of surviving. Thus, the maintenance of clean monocultures (i.e., cultures not contaminated with other bacteria) is facilitated. Conversely, to deactivate the bacteria, the reaction temperature is lowered to below the permissive  
25 range. The organisms cease growth, and hydrogen production stops. The organisms can be stored in a sealed bottle for several months with no harm or apparent loss of viability of the organisms. To kill these organisms they are sparged with air for 1-3 hours while the temperature is elevated to the permissive range

At temperatures in the permissive range, the bacteria convert a wide array of  
30 feedstocks into free hydrogen gas ( $H_2$ ) and lesser quantities of carbon dioxide ( $CO_2$ ). Only trace quantities of hydrogen sulfide ( $H_2S$ ) are generated if the concentrations of elemental sulfur and sulfide salts available to the bacteria in the medium and feedstock are sufficiently low.

Like the permissive range for temperature, there are "permissive ranges" for  
35 both pH and oxygen. The organisms function best at an optimal pH and oxygen level,

with diminishing functionality as the measured pH and oxygen levels differ from the optimal. The permissive range for both pH and oxygen levels also differs for each Species within the Order *Thermotogales*.

The pH at which the bacterial incubation process is conducted is extremely important, and greatly affects hydrogen gas yields. Titration or the addition of a physiological buffer to the incubation solution helps to maintain the pH and improves hydrogen yields. The optimal incubation pH (relative to optimal hydrogen production) is determined experimentally for each Genus and Species of bacteria in the Order *Thermotogales*. While the optimum pH will vary from Species to Species within the Order, a pH of between 4 and 10 is a good starting point to determine pH sensitivity. Preferably, pH specificity is determined empirically for each Species incorporated into the protocol. It should be noted that per convention, pH is measured at 20 °C, versus the actual running temperature.

Process Detail

A culture of bacteria of the Order *Thermotogales* is placed in a vessel along with a feedstock plus an aqueous medium. Under the incubation conditions tested, with over 3500 experiments conducted by the inventors, it was found that the bacteria produce virtually no nitrogen oxides, sulfur oxides, carbon monoxide, or methane gas. Reduced production of CO<sub>2</sub> and H<sub>2</sub>S is also obtained when elemental sulfur and sulfide salts are not available in the incubation medium.

Specifically, a culture of these bacteria is placed in a vessel along with an aqueous medium plus a feedstock. The aqueous medium contains a mixture of water, salts, vitamins, physiological buffers and biological cofactors. The feedstock is a material or waste that consists of or contains any combination of sugars (monosaccharides or polysaccharides) or complex hydrocarbons. At temperatures above 45°C and at lowered dissolved oxygen levels (reduced oxygen gas in the headspace gases relative to ambient air), the bacteria can convert a wide array of feedstocks into free hydrogen gas (H<sub>2</sub>) and lesser quantities of carbon dioxide (CO<sub>2</sub>). Only trace quantities of hydrogen sulfide (H<sub>2</sub>S) are generated if the sulfur (elemental or sulfide salts) content of the feedstock is sufficiently low. Hydrogen gas production does not appear to be end-product (H<sub>2</sub>) inhibited. To maintain continuous production, pH must be controlled by titration or with a physiological buffer. (A physiological buffer is one that acts to maintain a constant pH and is not toxic to living tissue.) This process does not require completely anaerobic conditions within the reactor, as in true fermentative processes

Prior to incubation and concomitant hydrogen production, nitrogen, or an inert

gas (which is not likely to be used by the organisms), is sparged into the aqueous medium to reduce the dissolved oxygen concentration in the aqueous medium to a level below that of water in equilibrium with ambient air. While the amount of oxygen in the aqueous medium is less than that normally found in water that is in equilibrium with 5 ambient air, some oxygen remains. During the sparging process, oxygen is also displaced from the headspace, thereby reducing the opportunity for reintroduction of oxygen into the aqueous medium. By capping or sealing the container immediately, oxygen is prevented from re-entering the bottle.

As an option, an indicator dye that changes color in the presence of elevated 10 concentrations of oxygen gas may be added to the aqueous medium. Examples of such indicator dyes include Resazurin or Methyl Green. However, once the appropriate amount of time has been empirically determined, the indicator dyes can be replaced by simply sparging for the amount of time empirically determined as adequate to reduce the oxygen content sufficiently.

15 The role of oxygen gas ( $O_2$ ) in the process of hydrogen gas generation has not yet been determined. However, it is reasonable to consider dissolved oxygen concentrations in terms of a "permissive range", which may differ with each Species. Generally, however, oxygen levels of between 0 and 15 volumetric percent of the gas in the head space of the reaction vessel is suitable.

20 In a batch culture, the maximum hydrogen gas concentration generated by most hydrocarbon sources occurs between 3-7 days from the start of incubation. The chief gases produced are hydrogen (as much as 25-35% volumetric percent of the head space gas), and carbon dioxide (approximately  $\frac{1}{2}$  the amount of hydrogen produced), and small quantities (ppm) of hydrogen sulfide.

25 In a continuously growing culture, it is theoretically possible that hydrogen product might reach as high as 60% with most of the balance being carbon dioxide. Initially nitrogen is needed to displace the oxygen from both the aqueous medium and the headspace; however, once sufficient hydrogen and carbon dioxide have evolved, the nitrogen sparge may no longer be needed. Hydrogen and carbon dioxide gases 30 would comprise almost the entire gas stream produced in a continuous production scheme.

In a continuously operating bio-reactor, the constituents of the aqueous medium and the headspace can be continuously or intermittently replenished, withdrawn or adjusted, as can the pH, percent dissolved oxygen, and temperature.

35 As noted supra, process temperatures of above 45 °C are suitable. With the

Species tested to date, the inventors have determined a preferable temperature range of approximately 55 °C and 90 °C.

Thermotogales

Order Detail

- 5 Either the literature or test results indicate that hydrogen production occurs with all nineteen readily available (and thus, testable) members (shown below) of the Order *Thermotogales*. This suggests that hydrogen production may be a defining characteristic of members of this entire Order. All members of the Order *Thermotogales* are considered suitable for use in the invented hydrogen producing process.
- 10 Several resources provide general protocols for establishing and maintaining viable cultures of bacteria of the Order *Thermotogales* for inoculation, including R. Huber et al. "The order Thermatogales" In The Prokaryotes, Eds. A Balows, et al (Springer, Berlin, Heidelberg, NY, 1992); incorporated herein by reference. The following members of the Order *Thermotogales* are suitable for use in the invented
- 15 hydrogen producing process:

*Thermotoga* sp. (DSMZ 4138)\*

*Thermotoga elfeii* (ATCC 51869, DSMZ 9442)

*Thermotoga hypogea* (DSMZ 11164)

*Thermotoga maritima* (DSMZ 3109)

- 20
- Thermotoga neapolitana* (from Naples) (ATCC 49049, DSMZ  
  4359)  
    *Thermotoga neapolitana* (from Africa) (DSMZ 5068)  
    *Thermotoga subterranea* (DSMZ 9912)  
    *Thermotoga thermarum* (DSMZ 5069)
- 25
- Petrotoga miotherma* (ATCC 51224, DSMZ 10691)  
  *Petrotoga mobilis* (DSMZ 10674)  
  *Thermosiphon* sp. (DSMZ 6568)  
  *Thermosiphon africanus* (DSMZ 5309)  
  *Thermosiphon melanesiensis* (DSMZ 6976)
- 30
- Fervidobacterium islandicum* (DSMZ 5733)  
  *Fervidobacterium nodosum* (DSMZ 5306)  
  *Fervidobacterium pennavorans* (DSMZ 9078)  
  *Fervidobacterium gondwanense* (DSMZ 13020)  
  *Geotoga petraea*
- 35
- Geotoga subterranea*

\*DSMZ = Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH  
(German Collection of Microorganisms and Cell Culture)

Nutrient and Media Detail

5 An aqueous medium comprising water, salts, vitamins, physiological buffers, and biological cofactors are utilized. The aqueous medium is supplemented with a hydrocarbon source as the "feedstock".

Bacteria of the Order *Thermotogales* can utilize very complex hydrocarbons without requiring preprocessing to produce simple sugars or their catabolites.

10 Therefore, feedstocks suitable for the invented process include materials, such as biomass or organic wastes, that consist of or contain any combination of: (1) sugars (monosaccharides and polysaccharides); (2) complex hydrocarbons, such as crude oil; (3) carbohydrates, such as starch; (4) xylans and celluloses; (5) amino acids; (6) proteins; and (7) long chain fatty acids.

15 Specific media formulations that have been demonstrated as suitable for specific members of the Order *Thermotogales* can be utilized in this process if the formulation is modified so that no elemental sulfur or sulfide salt is present. The elemental sulfur or sulfide salt is replaced with a compound such as cysteine, to provide any sulfur that might be needed by the bacteria. This modification will help minimize the production of

20 hydrogen sulfide.

Any media chosen is likely to be modified by enriching the hydrocarbon source, and/or omitting any indicator dyes (such as Resazurin or Methyl Green).

All of the common fermentative catabolites (such as acetate salts) are also omitted from the media formulations.

25 The following are exemplary media which can be employed in the process:

ATCC Medium 1658:	MMS Medium for <i>Thermotoga neap.</i>
ATCC Medium 1977:	<i>Thermotoga elfeii</i> Medium NH <sub>4</sub> Cl
ATCC Medium 1881:	<i>Petrotoga</i> Medium Trypticase
DSMZ Medium 88:	<i>Sulfolobus</i> Medium
DSMZ Medium 141:	<i>Methanogenium</i> Medium
DSMZ Medium 144:	<i>Thermoanaerobium</i> Medium
DSMZ Medium 343:	<i>Thermotoga</i> Medium
DSMZ Medium 483:	<i>Thermosiphon africanus</i> Medium
DSMZ Medium 498:	<i>Thermotoga II</i> Medium

	DSMZ Medium 501:	<i>Fervidobacterium islandicum</i> Medium
	DSMZ Medium 613:	TF Medium
	DSMZ Medium 644:	<i>Thermotoga elfeii</i> Medium
	DSMZ Medium 688:	<i>Thermotoga subterranea</i> Medium
5	DSMZ Medium 718:	<i>Petrotoga</i> Medium
	DSMZ Medium 740:	TF(A) Medium
	DSMZ Medium 794:	<i>Thermotoga hypogea</i> Medium

The following materials (see Demonstration Experiment) are likely to be found in a suitable medium for this process. Some of the constituents are consumed by the organisms and others are needed for the organism to prevent hyper- or hypo-tonicity and to maintain homeostasis. While exact concentrations of each constituent are not as important as its mere presence, the pH of the medium seems to require control to maintain high production rates.

Demonstration Experiment

- 15      The current known best practice protocol for preparation of the aqueous medium and for conducting a demonstration experiment is outlined below.
1.      An exemplary aqueous medium is prepared using the following constituents and protocol. It should be noted that different concentrations of salts should be used for some bacteria of the Order *Thermogales* that typically are found in an environment 20     that contains either more or less salt than in the illustrated examples contained herein.

Aqueous Medium

(Use a different concentration of salts for some species; see examples)

	H <sub>2</sub> O	1.0 L
	NH <sub>4</sub> Cl	1.0 gm
25	K <sub>2</sub> HPO <sub>4</sub>	0.3 gm
	KH <sub>2</sub> PO <sub>4</sub>	0.3 gm
	MgCl <sub>2</sub> x 6 H <sub>2</sub> O	0.2 gm
	CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.1 gm
	NaCl	10.0 gm
30	KCl	0.1 gm
	Cysteine HCl	1.0 gm
	Trizma Base (optional)	1.21 gm
	Sodium Acetate (optional)	0.5 gm

5

	Yeast Extract	2.0 gm
	Trypticase (BBL 11921)	2.0 gm
	Resazurin (optional)	0.5 mg
	Vitamin Solution (described below)	10.0 ml
	Trace Element Solution (described below)	10.0 ml

**Vitamin Solution**

(This solution can be omitted for some species; see examples)

	H <sub>2</sub> O	1.0 L
	Biotin	2.0 mg
10	Folic Acid	2.0 mg
	Pyridoxine HCl	10.0 mg
	Thiamine HCl	5.0 mg
	Riboflavin	5.0 mg
	Nicotinic Acid	5.0 mg
15	Calcium D-(+)-Pantothenate	5.0 mg
	Cyanocobalamin	0.1 µg
	p-Aminobenzoic Acid	5.0 mg
	Thioctic Acid	5.0 mg

**Trace Element Solution**

(This solution can be omitted for some species; see examples)

	Nitrolothriacetic acid	1.500 g
	MgSO <sub>4</sub> x 7 H <sub>2</sub> O	3.000 g
	MnSO <sub>4</sub> x 2 H <sub>2</sub> O	0.500 g
	NaCl	1.000 g
25	FeSO <sub>4</sub> x 7 H <sub>2</sub> O	0.100 g
	CoSO <sub>4</sub> x 7 H <sub>2</sub> O	0.180 g
	CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.100 g
	ZnSO <sub>4</sub> x 7 H <sub>2</sub> O	0.180 g
	CuSO <sub>4</sub> x 5H <sub>2</sub> O	0.010 g
30	KAl(SO <sub>4</sub> ) <sub>2</sub> x 12 H <sub>2</sub> O	0.020 g
	H <sub>3</sub> BO <sub>3</sub>	0.010 g
	Na <sub>2</sub> MoO <sub>4</sub> x 2H <sub>2</sub> O	0.010 g
	NiCl <sub>2</sub> x 6 H <sub>2</sub> O	0.025 g

Na<sub>2</sub>SeO<sub>3</sub> × 5 H<sub>2</sub>O  
Distilled water

0.300 mg  
1000.000 ml

2. The initial pH of the aqueous medium is adjusted with acid or base (e.g. HCl or  
5 NaOH) as appropriate for the selected species of bacteria.
3. In experimental situations, the aqueous medium is placed in a container that has  
an appropriate headspace and that can be sealed. An appropriate headspace is one  
that is large enough to collect the hydrogen gas. An example of a suitable container for  
10 bench-top analysis is a serum bottle with 150 ml capacity. In such situations,  
approximately 50 ml of medium is put in the bottle.
4. Approximately 0.25 grams of a feedstock are added to the reaction container.  
The feedstock contains one or more of the following: starch, cellulose, bagasse,  
15 carboxymethyl cellulose, hemicellulose, glucose, cellobiose, crude oil, amino acids,  
proteins or other hydrocarbon source.
5. Optionally, a physiological buffer (such as, Trizma Base) is added to the solution  
to help maintain a constant pH. Maintaining the pH of the cultured medium either with  
a physiological buffer or by titration facilitates optimal hydrogen gas production.
- 20 6. An indicator dye (e.g., Resazurin or Methyl Green) may be added as an optional  
indicator to show when a high concentration of dissolved oxygen is present. When  
sufficient O<sub>2</sub> is removed by the purging procedure discussed in step 7, the solution turns  
colorless.
7. Excess oxygen is removed from the reaction chamber by sparging with an inert  
25 gas for a time sufficient to reduce the oxygen content in both the liquid and gaseous  
phases in the container. Application of heat will facilitate the sparging process. While  
sparing, the reaction container is heated, (e.g., by being placed in a hot oil bath) at  
approximately 100°C for 5-10 minutes while the sparge gas is gently bubbled through  
the medium. If an indicator dye has been added, the sparging is stopped when the  
30 liquid medium goes colorless. Inasmuch as a standard serum bottle is utilized as the  
reaction vessel, sealing after sparging is nearly immediate. This helps prevent re-entry  
of oxygen into the reaction space.

8. The reaction chamber with its contents is then subjected to heat for a time sufficient to sterilize the contents. In this example, the chamber is sterilized at 134-138°C for ½ hour.
9. The solution is then inoculated with bacteria of the Order *Thermotogales*, an 5 example of which is *Thermotoga neapolitana* (Naples).
10. The bacteria are incubated for a time sufficient to produce hydrogen gas. Generally, suitable parameters include incubation times of between 3-9 days at a temperature of above 45 °C, preferably above 55 °C, and most preferably between 55 °C and 90 °C (with or without mixing) without elevating the pressure artificially.
- 10 11. Following incubation, gases in the headspace of the serum bottle, are withdrawn and analyzed using a hydrogen sensing device such as a gas chromatograph.
12. In a continuous fermentation, the hydrogen gas can be removed from the headspace and collected for analysis using a myriad of gas collection means, including gas-tight bags or containers.
- 15 This best practice demonstration experiment describes a basic process that can lead to the development of an industrial-scale production of hydrogen gas and carbon dioxide (2:1 v/v). The demonstration experiment starts with a liquor of 50 ml of the aqueous medium (as described above) and can produce at least 20 to 35 ml total hydrogen gas evolved in addition to the CO<sub>2</sub> and H<sub>2</sub>S in 3-7 days within a headspace 20 of 110 ml. This is accomplished using a biological process that degrades complex hydrocarbons. No other process has been found that describes the use of members of the Order *Thermotogales* in combination with the described mixture of salts, vitamins, physiological buffers and biological cofactors for such rapid production of hydrogen gas.
- 25 The following examples are illustrative only. As such, the specific concentrations and ranges discussed therein should not be construed as limiting the scope of the claims, appended hereto.

Example 1

The thermophilic bacteria, *Thermotoga neapolitana* (ATCC 49049, DSMZ4359 from Naples) was placed in a sealed container along with the medium described above, at 30 the strength indicated above, pH 8.5, and with soluble starch as the hydrocarbon

source. The container had 50 ml of liquor with a headspace of 110 ml, consisting initially of nitrogen gas with 5-12% oxygen gas. The hydrogen gas production in the headspace may be described as follows: (1) a cumulative H<sub>2</sub> production of 24.4 ml (in 110 ml of available headspace) in 85 hours of incubation; (2) a cumulative H<sub>2</sub> production rate of 2.61 ml/l-hour; and a calculated H<sub>2</sub> concentration of 221 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 2

A thermophilic bacteria, *Thermotoga neapolitana* (ATCC 49049, DSMZ 4359 from Naples), was placed in a sealed container along with a physiologically buffered solution of the medium described above, at the concentration noted, with a pH of 8.0, and with soluble starch as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of at least 33.7 ml (in 110 ml of available headspace) in 74 hours; (2) a cumulative H<sub>2</sub> production rate of at least 4.14 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 306 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 3

A thermophilic bacteria, *Petrotoga mobilis* (DSMZ 10674), was placed in a sealed container along with the medium described above at double strength with a pH of 6.0 and cellobiose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 15.5 ml (in 110 ml of available headspace) in 248 hours; (2) a cumulative H<sub>2</sub> production rate of .567 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 140 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 4

A thermophilic bacteria, *Thermotoga elfeii* (ATCC 51869, DSMZ 9442), was placed in a sealed container along with a buffered solution of the medium described above at the concentration noted, with a pH of 8.0, and with soluble starch as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 31.6 ml (in 110 ml of available headspace) in 124 hours; (2) a cumulative H<sub>2</sub> production rate of at least 2.32 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 287 ml/l. This production

assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 5

A thermophilic bacteria, *Petrotoga miotherma* (ATCC 51224, DSMZ 10691), was

- 5 placed in a sealed container along with the unbuffered medium described above at a pH of 8.0 and with potato starch as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 12.9 ml (in 110 ml of available headspace) in 176 hours; (2) a cumulative H<sub>2</sub> production rate of 0.668 ml/l-hour; and  
10 (3) a calculated H<sub>2</sub> concentration of 118 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 6

A thermophilic bacteria, *Thermotoga* sp.(DSMZ 4138), was placed in a sealed container

- 15 along with the medium described above at a pH of 8.0, and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of at least .56 ml (in 110 ml of available headspace) in 55 hours; (2) a cumulative H<sub>2</sub> production rate of at least 0.093 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 50.9  
20 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 7

A combination of thermophilic bacteria, *Thermotoga eifelii* (ATCC 51869, DSMZ 9442), *Thermotoga neapolitana* (ATCC 49049, DSMZ 4359) and *Thermotoga* sp. (DSMZ

- 25 4138), were placed in a sealed container along with a double strength medium compared to that described above, at a pH of 6.5 and with cellulose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 18.0 ml (in 110 ml of available headspace) in 146 hours; (2) a cumulative H<sub>2</sub> production rate of at  
30 least 1.12 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 163 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 8

- The thermophilic bacteria, *Thermotoga hypogea* (DSMZ 11164) was placed in a sealed container along with a single strength medium compared as described above, at a pH of 9.0 and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) 5 a cumulative H<sub>2</sub> production of 20.08 ml (in 110 ml of available headspace) in 179 hours; (2) a cumulative H<sub>2</sub> production rate of at least 1.02ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 182.5 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 9

- 10 The thermophilic bacteria, *Thermosiphon africanus* (DSMZ 5309) was placed in a sealed container along with a double strength medium compared with the media described above, at a pH of 8.0 and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 21.99 ml (in 110 ml of available headspace) in 111 hours; (2) a cumulative H<sub>2</sub> production rate of at least 1.80 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 199.9 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 10

- 20 The thermophilic bacteria, *Fervidobacterium pennavorans* (DSMZ 9078) was placed in a sealed container along with a one-tenth strength medium compared with the medium described above, at a pH of 8.0 and with cellobiose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 17.68 ml (in 110 ml of available headspace) in 80 hours; (2) a cumulative H<sub>2</sub> production rate of at least 2.01 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 160.7 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 11

- 30 The thermophilic bacteria, *Thermotoga subterranea* (DSMZ 9912) was placed in a sealed container along with DSMZ medium 344, a medium similar to that described in detail above, at a pH of 7.0 and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub> production of 19.49 ml (in 110 ml of available

headspace) in 128 hours; (2) a cumulative H<sub>2</sub> production rate of at least 1.38 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 177.2 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

5    Example 12

The thermophilic bacteria, *Thermotoga maritima* (DSMZ 3109) was placed in a sealed container along with a single strength DSMZ medium 688, a medium similar to that described above, at a pH of 7.0 and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production can be  
10 described as follows: (1) a cumulative H<sub>2</sub> production of 6.31 ml (in 110 ml of available headspace) in 128 hours; (2) a cumulative H<sub>2</sub> production rate of at least 0.448 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 57.4 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

15    Example 13

The thermophilic bacteria, *Thermosiphon melanesiensis* (DSMZ 12029) was placed in a sealed container along with single strength DSMZ medium 344, a medium similar to that described in detail above, at a pH of 7.0 and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas production  
20 can be described as follows: (1) a cumulative H<sub>2</sub> production of 17.97 ml (in 110 ml of available headspace) in 128 hours; (2) a cumulative H<sub>2</sub> production rate of at least 1.28 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 163.4 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

25    Example 14

The thermophilic bacteria, *Thermotoga neapolitana* (africa) (DSMZ 5068) was placed in a sealed container along with a single strength DSMZ medium 344, a medium similar to the medium described above, at a pH of 7.0 and with dextrose as the hydrocarbon source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas  
30 production can be described as follows: (1) a cumulative H<sub>2</sub> production of 13.63 ml (in 110 ml of available headspace) in 175 hours; (2) a cumulative H<sub>2</sub> production rate of at least 0.708 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 123.9 ml/l. This

production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

Example 15

The thermophilic bacteria, *Fervidobacterium gondwanense* (DSMZ 13020) was placed  
5 in a sealed container along with a single strength DSMZ medium 740, a medium similar  
to the medium described above, at a pH of 7.0 and with dextrose as the hydrocarbon  
source. There was 50 ml of liquor with a headspace of 110 ml. The hydrogen gas  
production can be described as follows: (1) a cumulative H<sub>2</sub> production of 6.57ml (in  
110 ml of available headspace) in 14 hours; (2) a cumulative H<sub>2</sub> production rate of at  
10 least 5.4 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of 59.72 ml/l. This production  
assumes that the hydrocarbon source was not rate-limiting and that the tests started  
when the sterile medium was inoculated.

Example 16

The thermophilic bacteria, *Thermotoga neapolitana* (DSMZ 4359 from Naples) was  
15 placed in a sealed container along with ATCC medium 1977 and no additional  
hydrocarbon source was added beyond the proteins, amino acids and hydrocarbons  
characteristic of this medium. The bacteria had previously been habituated to a medium  
containing only minimal amounts of carbohydrates. There was 50 ml of liquor with a  
headspace of 110 ml in the container. The hydrogen gas production can be described  
20 as follows: (1) a cumulative H<sub>2</sub> production of 5.12ml (in 110 ml of available headspace)  
in 137 hours; (2) a cumulative H<sub>2</sub> production rate of at least .339 ml/l-hour; and (3) a  
calculated H<sub>2</sub> concentration of 46.54 ml/l. This production assumes that the  
hydrocarbon source was not rate-limiting and that the tests started when the sterile  
medium was inoculated.

25 Example 17

The thermophilic bacteria, *Thermotoga elfeii* (ATCC 51869, DSMZ 9442) was placed in  
a sealed container along with ATCC medium 1977 at a pH of 8.0 and with West Virginia  
Crude Oil as the hydrocarbon source. There was 50 ml of liquor with a headspace of  
110 ml. The hydrogen gas production can be described as follows: (1) a cumulative H<sub>2</sub>  
30 production of 11.20 ml (in 110 ml of available headspace) in 183 hours; (2) a cumulative  
H<sub>2</sub> production rate of at least .556 ml/l-hour; and (3) a calculated H<sub>2</sub> concentration of  
101.80 ml/l. This production assumes that the hydrocarbon source was not rate-limiting  
and that the tests started when the sterile medium was inoculated.

Example 18

The thermophilic bacteria, *Thermosiphon* sp. (DSMZ 6568) was placed in a sealed container along with DSMZ medium 613, a medium similar to the example medium above, and listed as one of the media that can be used for this invention. A pH of 7.0  
5 was used glucose as the hydrocarbon source. The container had 50 ml of liquor with a headspace of 110 ml, consisting initially of nitrogen gas with 5-12% oxygen gas. The hydrogen gas production in the headspace may be described as follows: (1) a cumulative H<sub>2</sub> production of 13.1 ml (in 110 ml of available headspace) in 22 hours of incubation; (2) a cumulative H<sub>2</sub> production rate of 5.4 ml/l-hour; and a calculated H<sub>2</sub>  
10 concentration of 119 ml/l. This production assumes that the hydrocarbon source was not rate-limiting and that the tests started when the sterile medium was inoculated.

While the invention has been described with reference to details of the illustrated embodiment, these details are not intended to limit the scope of the invention as defined in the appended claims. For example, many species in the Order *Thermotogales* have  
15 been isolated from oil wells. Inasmuch as these organisms appear to be able to live and proliferate within the oil wells, then these organisms should be able to consume the organic matter found there, to produce the large amounts of H<sub>2</sub>S normally found at those depths. In fact, the inventors have found that these bacteria produce increased amounts of H<sub>2</sub>S when fed crude oil.

20 In light of the foregoing, a method for producing hydrogen is proposed wherein organisms from the Order *Thermotogales* are injected into tertiary oil wells that have been pre-treated in a manner that sequesters sulfides and elemental sulfur. Care is taken to maintain suitable pH, salinity, and temperature and perhaps pressure, of the spent well-turned reaction vessel. The envisioned process would facilitate consumption  
25 of residual oil which is no longer economically feasible for extraction. Harvesting procedures of the resulting hydrogen gas is effected via conventional means, i.e., vacuum, or sweeping with inert fluids.

Inasmuch as the oil-feedstocks in these scenarios are several thousand feet below the earth's surface, any inhibiting effect due to the oxygen concentrations found  
30 at the earth's surface (i.e., 20 percent) is nonexistent.

## CLAIMS

The embodiment of the invention in which an exclusive property or privilege is claimed is defined as follows:

1. A process for producing H<sub>2</sub> comprising:
  - a) confining bacteria from the Order *Thermotogales* to an environment suitable for allowing the bacteria to undergo metabolism;
  - b) maintaining the environment at an oxygen concentration below the oxygen concentration of water in equilibrium with air; and
  - c) removing hydrogen from the environment during the metabolism.
2. The process as recited in claim 1 wherein the environment has no added S<sup>0</sup> or inorganic sulfide salts.
3. The process of claim 1 wherein the temperature of the environment is maintained within the permissive range of temperatures for hydrogen production by the Species of bacteria in use.
4. The process as recited in claim 1 wherein the environment is maintained at a temperature above 45 °C.
5. The process as recited in claim 1 wherein the environment contains a feedstock selected from the group consisting of simple sugars, starches, xylans, celluloses, oils, petroleums, bitumens, amino acids, long-chain fatty acids, proteins and combinations thereof.
6. The process of claim 1 wherein the oxygen gas (O<sub>2</sub>) concentration of the environment, either dissolved within an aqueous medium or in a gaseous state, is

maintained within the permissive range of oxygen gas concentrations for hydrogen gas production by the Species of bacteria in use.

7. The process as recited in claim 1 wherein oxygen gas concentration in the environment decreases with the metabolism.

8. The process as recited in claim 1 wherein oxygen gas concentration in the environment increases with the metabolism.

9. The process as recited in claim 1 wherein the environment is maintained at between 55 °C and 90 °C.

10. The process of claim 1 wherein the pH of the environment is maintained within the permissive range of pH for the Species of bacteria in use.

11. The process as recited in claim 1 wherein the environment is maintained at a pH of between approximately 4 and 10.

12. The process as recited in claim 1 wherein the bacteria is a species selected from the group consisting of *Thermotoga neapolitana* J., *Petrotoga mithoferma*, *Petrotoga mobilis*, *Thermotoga* sp., *Thermotoga eletii*, *Thermotoga hypogea*, *Thermosiphon africanus*, *Fervidobacterium pennavorans*, *Thermotoga neapolitana*, *Thermotoga subterranea*, *Fervidobacterium gondwanense*, *Thermosiphon melanesiensis*, *Thermotoga maritima*, *Thermotoga thermarum*, *Thermosiphon* sp., *Fervidobacterium islandicum*, *Fervidobacterium nodosum*, *Geotoga petraea*, or *Geotoga subterranea*, and combinations thereof.

13. The process as recited in claim 1 wherein the process is optimized to yield rates that may exceed 5 ml/l-hour.

14. A method for producing H<sub>2</sub> comprising:

- a) selecting a bacteria from the Order *Thermotogales*;
- b) subjecting the bacteria to an environment containing a feedstock and to a media, wherein the media contains oxygen gas in a concentration below the oxygen concentration of water in equilibrium with air; and
- c) maintaining the environment at a temperature, pressure and pH sufficient to allow the bacteria to metabolize the feedstock.

15. The process as recited in claim 14 wherein the oxygen gas ( $O_2$ ) concentration of the environment, either dissolved within an aqueous medium or in a gaseous state, is maintained within the permissive range of oxygen gas concentrations for hydrogen gas production by the Species of bacteria in use..

16. The method as recited in claim 14 wherein the oxygen concentration decreases as the metabolism occurs.

17. The method as recited in claim 14 wherein the oxygen concentration increases as the metabolism occurs.

18. The process as recited in claim 14 wherein the bacteria is a species selected from the group consisting of *Thermotoga neapolitana* J., *Petrotoga miotherma*, *Petrotoga mobilis*, *Thermotoga* sp., *Thermotoga elfeii*, *Thermotoga hypogea*, *Thermosiphon africanus*, *Fervidobacterium pennavorans*, *Thermotoga neapolitana*, *Thermotoga subterranea*, *Fervidobacterium gondwanense*, *Thermosiphon melanesiensis*, *Thermotoga maritima*, *Thermotoga thermarum*, *Thermosiphon* sp., *Fervidobacterium islandicum*, *Fervidobacterium nodosum*, *Geotoga petraea*, or *Geotoga subterranea*, and combinations thereof.

19. The method as recited in claim 14, wherein the feedstock is a sugar selected from the group consisting of simple sugars, starches, xylans, celluloses, and combinations thereof.

20. The method as recited in claim 14, wherein the feedstock is a complex hydrocarbon selected from the group consisting of oils, petroleums, bitumens, amino acids, long-chain fatty acids, proteins, and combinations thereof.

21. The process as recited in claim 14 wherein the temperature of the environment is maintained within the permissive range of temperatures for hydrogen production by the Species of bacteria in use.

22. The method as recited in claim 14, wherein the environment is maintained at a temperature above 45° C.

23. The method as recited in claim 14, wherein the suitable growth environment is maintained at a temperature of between 55 °C and 90 °C.

24. The process as recited in claim 14 wherein the pH of the environment is maintained within the permissive range of pH for the Species of bacteria in use.

25. The method as recited in claim 14, wherein the environment is maintained at a pH of between approximately 4 and 10.

26. The method as recited in claim 14, wherein the environment is preconditioned with an inert gas selected from the group consisting of helium, neon, argon, krypton, xenon, and combinations thereof.

27. The method as recited in claim 14 wherein the media has no added S<sup>0</sup> or inorganic sulfide salts.

28. The method as recited in claim 14 wherein the aqueous medium contains an indicator for oxygen gas presence.

29. The method as recited in claim 14 wherein nitrogen is used to initially or continuously sparge or purge oxygen gas (O<sub>2</sub>) from the aqueous medium or headspace, with or without the assistance of a vacuum.

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(54) Title: PROCESS FOR GENERATION OF HYDROGEN GAS FROM VARIOUS FEEDSTOCKS USING THERMOPHILIC BACTERIA

(57) Abstract: A method for producing hydrogen gas is provided comprising selecting a bacteria from the Order Thermotogales, subjecting the bacteria to a feedstock and to a suitable growth environment having an oxygen concentration below the oxygen concentration of water in equilibrium with air, and maintaining the environment at the predetermined pH and at a temperature of at least approximately 45°C for a time sufficient to allow the bacteria to metabolize the feedstock.

## INTERNATIONAL SEARCH REPORT

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## B. FIELDS SEARCHED

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Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, BIOSIS, FSTA, MEDLINE, CHEM ABS Data

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 268 280 A (STARNES ROBERT L ET AL) 7 December 1993 (1993-12-07) column 4, line 43 - line 62; example 1 ---	1-29
X	ADAMS M W W: "Hydrogen from hot bacteria." CHEMTECH, vol. 11, 1991, pages 692-699, XP008006324 cited in the application the whole document ---	1-29
A	US 5 804 424 A (KAPLAN SAMUEL ET AL) 8 September 1998 (1998-09-08) cited in the application example 9 ---	29 -/-

Further documents are listed in the continuation of box C.

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## INTERNATIONAL SEARCH REPORT

In Application No  
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## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 4 921 800 A (VATSALA T M) 1 May 1990 (1990-05-01) cited in the application column 1, line 46 - line 53; claims 14,15 -----	26
A	US 5 350 692 A (TAGUCHI FUMIAKI ET AL) 27 September 1994 (1994-09-27) cited in the application -----	
A	US 5 834 264 A (BREZNAK JOHN A ET AL) 10 November 1998 (1998-11-10) cited in the application -----	
T	VAN OOTEGHEM, SUELLEN A. ET AL: "Hydrogen production by the thermophilic bacterium Thermotoga neapolitana" APPLIED BIOCHEMISTRY AND BIOTECHNOLOGY (2002), 98-100(BIOTECHNOLOGY FOR FUELS AND CHEMICALS), 177-189 , XP008006346 -----	

## INTERNATIONAL SEARCH REPORT

Information on patent family members

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